To TOSHIBA CARRIER (THAILAND) CO., LTD.

Test Report

Evaluation test of the removal efficacy of the Room air conditioner on airborne bacteria

(25 m³ space)

KRCES - Bio. Test Report No.2018_0357 February 15, 2019

Approved by: Tashihir Itoh

Toshihiro Itoh, Chief Director

Kitasato Research Center for Environmental Science
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The contents of this report should not be disclosed to the public without prior consent of the Kitasato Research Center for Environmental Science. The test results shown here are applied to only test samples and do not guarantee quality of the whole batch (lot) of the test material.

To TOSHIBA CARRIER (THAILAND) CO., LTD.

Test Report

Evaluation test of the removal efficacy of the Room air conditioner on airborne virus $(25 \text{ m}^3 \text{ space})$

KRCES - Bio. Test Report No.2018_0358 February 15, 2019

Toshihiro Ito

Approved by:

Toshihiro Itoh, Chief Director

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1. Test objective

Removal efficacy of the room air conditioners on airborne virus in a 25 m³ test chamber was evaluated in this study. Evaluation method for removal efficacy was conducted in accordance with Annex D, "The evaluation test for removal efficacy on airborne virus" in Japan Electrical Manufacturers' Association standard JEM 1467 "household air cleaner".

2. Client

Name: TOSHIBA CARRIER (THAILAND) CO., LTD.

Address: 144/9 Moo 5, Bangkadi, Industrial Park, Tivanon Road, Tambol Bangkadi, Amphur, Muang, Pathunthani 12000, Thailand

3. Test laboratory

Name: Kitasato Research Center for Environmental Science

Address: 1-15-1 Kitasato, Minami, Sagamihara, Kanagawa 252-0329, Japan

4. Test period

January 16, 2019 ~ January 18, 2019

5. Test sample

- 1) Room air conditioner A (Model No; RAS-35PKVPG-ND, Device; DSK9 with plasma) · · · Photo1
- 2) Room air conditioner B (Model No; RAS-13J2KCVRG-T, Device; BF1-13K with ionizer) · · · Photo2

6. Test condition

I. Natural reduction (control);

Time-dependent changes in virus count were monitored after the test virus suspension was splayed into the chamber where the test sample was turned off.

II. Room air conditioner A (The air blower only)

Time-dependent changes in virus count were monitored after the test virus suspension was sprayed into the chamber where only the air blower of the test sample was turned on.

III. Room air conditioner A (The air blower and the test device)

Time-dependent changes in virus count were monitored after the test virus suspension was sprayed into the chamber where the sample (the air blower and the test device) was turned on.

IV. Room air conditioner B (The air blower only)

Time-dependent changes in virus count were monitored after the test virus suspension was sprayed into the chamber where only the air blower of the test sample was turned on.

V. Room air conditioner B (Continuous operation of the air blower and the test device)

Time-dependent changes in virus count were monitored after the test virus suspension was sprayed into the chamber where the sample (the air blower and the test device) was continuously operated for 30 seconds, and then turn off for 2 seconds.

VI. Room air conditioner B (Intermittent operation of the air blower and the test device)

Time-dependent changes in virus count were monitored after the test virus suspension was sprayed into the chamber where the sample (the air blower and the test device) was intermittently operated for 4 seconds, and then turn off for 6 seconds.

7. Test microorganisms

Virus: Escherichia coli phage φX-174 NBRC 103405 (=ATCC 13706-B1)

Size: approximately 30 nm

Host bacteria: Escherichia coli NBRC 13898 (= ATCC 13706)

8. Reagents, apparatus

- 1) Main reagents
 - Nutrient broth (Difco)
 - · Sodium chloride (Wako, special grade, for physiological saline)
 - · Agar (Difco)
 - · Nutrient agar (Nissui)
 - Phosphate buffered saline (Elmex)
 - · Sodium thiosulfate (Wako, 1st grade)
- 2) Main apparatus
 - Test chamber (25 m³: $2.7 \times 3.8 \times 2.4$ m, Amenity Technology)

- · Circulation fan (BS-B-25, Yamazen)
- · Laser particle counter (MODEL 3886, Kanomax Japan)
- Thermo-hygrometer (TR-72Ui, T&D)
- · Nebulizer (Collison Nebulizer CN-31I、BGI)
- · Glass impinger (specially ordered)
- Membrane filter (ϕ 0.22 μ m, Bottle Top Filter, TPP)
- · Incubator (MIR-15 MIR-553, Sanyo)

9. Method

1) Test procedure

The test system was shown in Photos 3, 4 and Figs 5, 6. The test sample, the circulation fan, the laser particle counter, and the thermo-hygrometer were set in the test chamber. Two holes were made on one of the side panels of the test chamber. The nebulizer for spraying virus suspension was connected to one of the holes and the impinger for collecting airborne virus was connected to the other hole.

According to the test procedure described in Table 6, the virus suspension was sprayed with nebulizer for 15 minutes into the chamber while the circulation fan was operated. After 2 minute circulation of the air, the virus aerosol was collected into the impinger (time 0). The aerosol was collected after 60 and 120 minutes after the test sample was turned on.

As a control, the same test was performed under the condition that the test sample was turned off according to the procedure described in Table 5.

2) Preparation of test virus

The test virus was inoculated into the host bacterial suspension in nutrient broth which had been incubated at 36 ± 2 °C overnight. The virus/bacterial suspension was mixed with the semisolid agar medium (nutrient broth + 0.5% sodium chloride + 0.5% agar) and poured onto top of the agar medium. After 18 hour incubation at 36 ± 2 °C, the culture was centrifuged and the resultant supernatant was filtrated through a 0.22 µm membrane filter to remove the host bacteria, and 10^{10} PFU/mL of virus was isolated. The virus suspension was diluted 10-fold with sterile ion-exchanged water for the test.

3) Spray of the virus suspension

The test virus suspension was sprayed into the test chamber with the nebulizer for 15 minutes at the liquid rate of 0.2 mL/min. The air pressure from the compressor discharge was set at 1.5 kg/cm² and the air flow rate was set to 7.0 L/min.

4) Collection of airborne virus

The virus aerosol in the chamber was collected at 10 L/min for 5 minutes (total 50 L) to the glass impinger containing 20 mL of phosphate buffered saline added with 0.015% sodium thiosulfate.

5) Count of virus number

Ten-fold dilutions of the each collected virus suspension were prepared with phosphate buffered saline. The each dilution was mixed with *E.coli*, and spread on nutrient agar with the semisolid agar, and incubated at 36 ± 2 °C for 18 hours. Plaques were counted to calculate the numbers of virus in 50 L of air was calculated.

6) Evaluation method for the removal efficacy

This test was carried out using Annex D of JEM 1467 as a reference. In JEM 1467, achieving 2.0-digit reduction in 90 minutes in the test space of $20\sim32$ m³ is required to conclude that the test sample is effective.

However, this test sample did not correspond to household air purifiers. Therefore, the evaluation of the removal efficacy was performed using the method described below.

The approximate equation was calculated based on the time-dependent changes of airborne virus (logarithmic representation) and the inclination of the approximate equation was obtained. This inclination represents an amount of change in the number of virus per minute. Net inclination^{*1} was calculated by subtracting the inclination of control condition from that of test condition. Net LRV^{*2} and percent reduction^{*3} were calculated from the value of net inclination, and the removal efficacy of airborne virus was determined based on net LRV.

The efficacy of the test sample was evaluated using the following formulae.

*1 Net inclination =

Inclination of test sample condition — Inclination of control condition *2 Net LRV = -{ Net inclination \times Test time (min) }

**3 Percent reduction(%) =
$$\left(1 - \frac{1}{10^{\text{(Net LRV)}}}\right) \times 100 \text{ (%)}$$

10. Results

Virus count of the sprayed suspension was 5.5×10^8 PFU/mL. The numbers of airborne virus at each measurement time were shown in Table 1 and Fig 1. Net LRV and percent of reduction were calculated from the number of airborne virus at each measurement time, and were shown in Table 2 and Fig 2. Residual virus ratios for the initial number at each time were shown in Table 3 and Fig 3. The residual virus ratios for the control were shown in Table 4 and Fig 4.

In this test, net LRV (reduction rate) of the test samples for airborne virus at 120 minutes were -0.17 (-47%) for test condition II, 0.81 (84%) for the condition III, 0.04 (8%) for the condition IV, 1.40 (96%) for the condition V and 0.92 (87%) for the condition VI respectively.

11. Reference data

The number of airborne particles, temperature and humidity in the test chamber were shown as a reference data.

12. Comment

This test was referring to the method of JEM 1467. In attached document Annex D, "The evaluation test for removal efficacy on airborne virus" in Japan Electrical Manufacturers' Association standard JEM 1467, achieving 2.0-digit reduction in 90 minutes in the test space of $20\sim32$ m³ is required to conclude that the test sample is effective.

In this study, the test sample showed certain degree of reduction effect but could not achieved 2.0 log reduction value even after being operated for 120min.

The difference between this standard and this test must be considered, as follows.

• The test samples in this test were not designed as air purifiers but air conditioners.

Table 1. The number of airborne virus at each measurement time (Unit: CFU/50 L-air)

Test condition	Time (min)				
Test condition	0	60	120		
I. Control condition	400,000	53,000	40,000		
II . Room air conditioner A	160,000	21.000	94.000		
(The air blower only)	160,000	31,000	24,000		
III. Room air conditioner A	200,000	99,000	E 000		
(The air blower and the test device)	380,000	22,000	5,800		
IV. Room air conditioner B	220,000	20,000	20,000		
(The air blower only)	330,000	36,000	30,000		
V. Room air conditioner B	010 000	01.000	2 200		
(Continuous operation of the air blower and the test device)	810,000	21,000	3,200		
VI. Room air conditioner B	460,000	96,000	E 500		
(Intermittent operation of the air blower and the test device)	460,000	26,000	5,500		

Test sample:

- 1) Room air conditioner A (Model No; RAS-35PKVPG-ND, Device; DSK9 with plasma)
- 2) Room air conditioner B (Model No; RAS-13J2KCVRG-T, Device; BF1-13K with ionizer)

Test virus : *Escherichia coli* phage φX-174 NBRC 103405 (=ATCC 13706-B1)

Test space: 25 m³

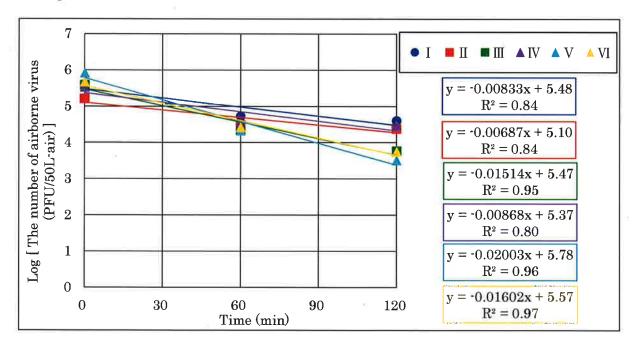


Fig 1. The number of airborne virus at each measurement time

Table 2. Net Litt and percent reduction at each measurement time	Table 2.	Net LRV an	d percent reduction	n at each measurement tim	1e
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The state of 1:4:	T1:4:	Net	7	Time (min))
Test condition	Inclination	inclination	0	60	120
I. Control condition	-0.00833				
II. Room air conditioner A	-0.00687	0.00146	0.00	-0.08	-0.17
(The air blower only)			(0%)	(-20%)	(-47%)
III. Room air conditioner A	-0.01514	-0.00681	0.00	0.40	0.81
(The air blower and the test device)			(0%)	(60%)	(84%)
IV. Room air conditioner B	-0.00868	-0.00035	0.00	0.02	0.04
(The air blower only)			(0%)	(4%)	(8%)
V. Room air conditioner B	0.00000	0.01150	0.00	0.70	1.40
(Continuous operation of the air blower and the test device)	-0.02003	-0.01170	(0%)	(80%)	(96%)
VI. Room air conditioner B			0.00	0.46	0.92
(Intermittent operation of the air blower and the test device)	-0.01602	-0.00769	(0%)	(65%)	(87%)

Net inclination = Inclination of test condition - Inclination of control condition

 $Net \; LRV \; = \; - \{ \; Net \; inclination \; \times \; Test \; time \; \; (min) \; \; \}$

Percent reduction (%) =
$$\left(1 - \frac{1}{10^{\text{(Net LRV)}}}\right) \times 100 \text{ (%)}$$

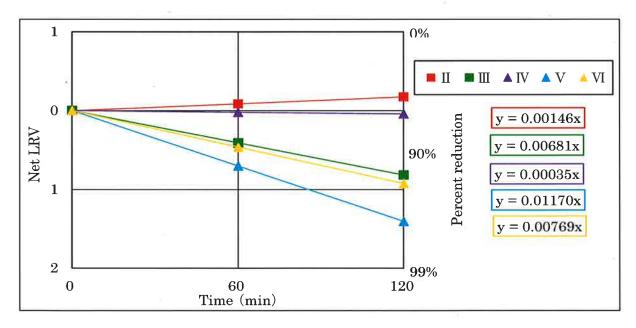


Fig 2. Net LRV and percent reduction at each measurement time

Table 3. R	esidual viru	s ratio for the	e initial number	(%	at each time
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Tract condition			Ti	me (mi	n)		
Test condition		20	40	60	80	100	120
I. Control condition	100	68	46	31	21	14	10
II. Room air conditioner A	100	70	E 0	20	00	90	1.4
(The air blower only)	100	72	5 3	38	28	20	14
III. Room air conditioner A	100	49	24	10	6	3.0	1 5
(The air blower and the test device)	100	49	24	12	0	3.0	1.5
IV. Room air conditioner B	100	67	44	30	20	13	9
(The air blower only)	100	07	44	30	20	10	9
V. Room air conditioner B							
(Continuous operation of the air blower and	100	39	15	6	2.4	0.9	0.3
the test device)							
VI. Room air conditioner B							
(Intermittent operation of the air blower	100	47	22	10	5	2.5	1.1
and the test device)							

*Using slope of approximate expression as shown in Fig1 (number of airborne virus which change per minute, log value) the residual ratio was calculated according to equation ① and ②.

① Log reduction value = | Inclination | ×Time (min)

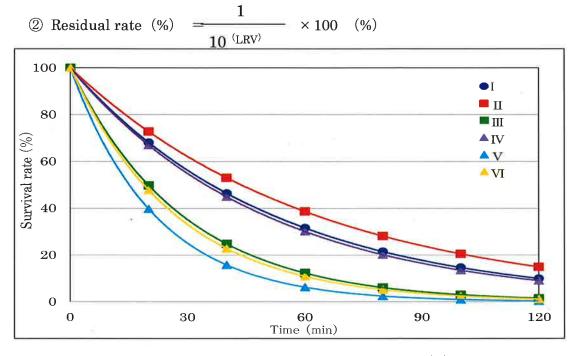


Fig 3. Residual virus ratio for the initial number (%) at each time

Table 4. The residual virus ratios with the sample for the control at each time

Toot andition			Ti	me (mi	n)		
Test condition		20	40	60	80	100	120
I. Control condition	100	100	100	100	100	100	100
II . Room air conditioner A (The air blower only)	100	106	114	122	130	139	149
III. Room air conditioner A (The air blower and the test device)	100	73	5 3	39	28	20	15
IV. Room air conditioner B (The air blower only)	100	98	96	95	93	92	90
V. Room air conditioner B (Continuous operation of the air blower and the test device)	100	58	34	19	11	6.7	3.9
VI. Room air conditioner B (Intermittent operation of the air blower and the test device)	100	70	49	34	24	17	11

*Using slope of approximate expression as shown in Fig1 (number of airborne virus which change per minute, log value) the residual ratio was calculated according to equation ①,② and ③.

- ① Net inclination = Inclination of test condition Inclination of control condition
- ② LRV= | Net Inclination | ×Time (min)

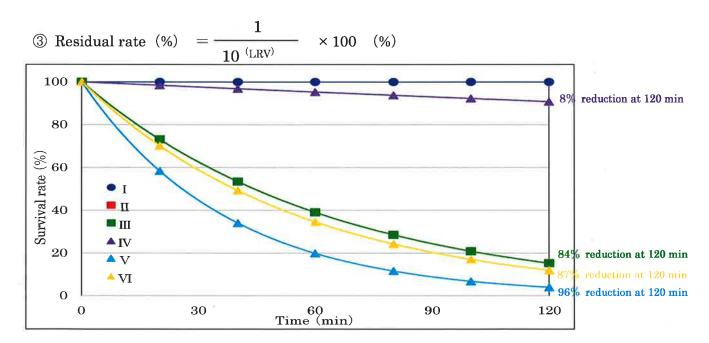


Fig 4. The residual virus ratios with the sample for the control at each time

Table 5. Test procedure (Control condition I)

Test condition	Equipment		Time (mi	n)	
Test condition	Equipment	0	6	0	120
To make homogeneous air in the chamber	Circulation fan			-	-
Spray of virus	Nebulizer	15min Dispersi	ng bacteria for 2	min	
Collect airborne virus	Impinger	5min 10L/min	_	5min 10L/min	5min 10L/min

Table 6. Test procedure (Test condition $\text{II} \sim \text{VI}$)

Test condition	Equipment		Time (mi	n)	
Test condition	Equipment	0	6	0 1	20
To make homogeneous air in the chamber	Circulation fan				-
Spray of virus	Nebulizer	15min Dispersin	ng bacteria for 2 i	min	
Test sample	Air blower				\rightarrow
rest sample	Test device		/		
Collect airborne virus	Impinger	5min 10L/min		5min 10L/min	5min 10L/min



Photo 1. Room air conditioner A (Model No ; RAS-35PKVPG-ND)



Photo 2. Room air conditioner B (Model No; RAS-13J2KCVRG-T)



Photo 3. 25 m³ Test chamber (Room air conditioner A)



Photo 4. 25 m³ Test chamber (Room air conditioner B)

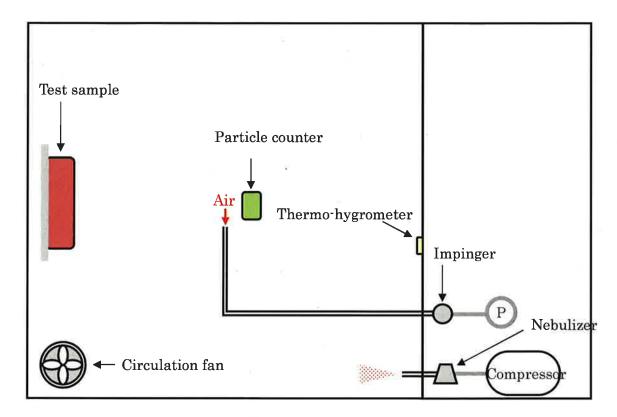


Fig 5. 25 m³ Test chamber (top view)

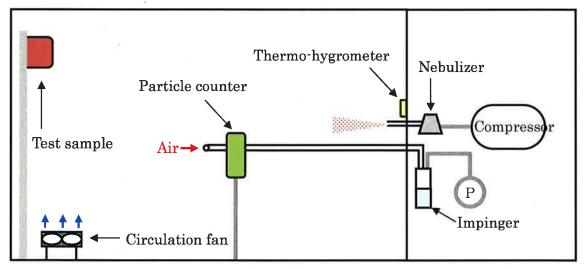
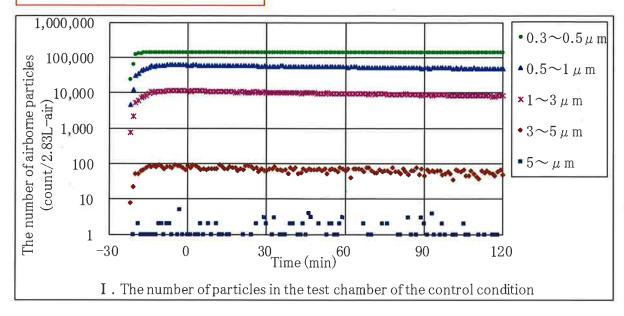
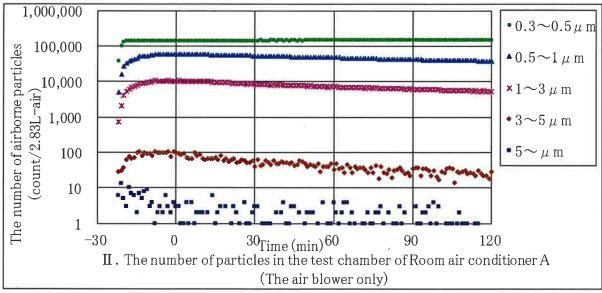
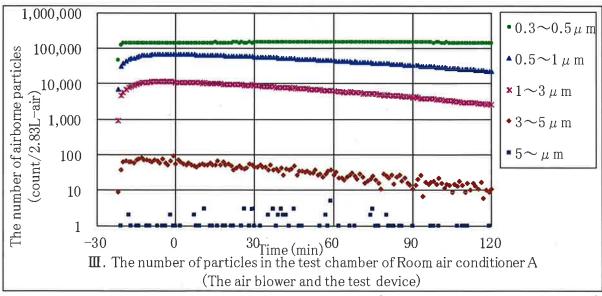


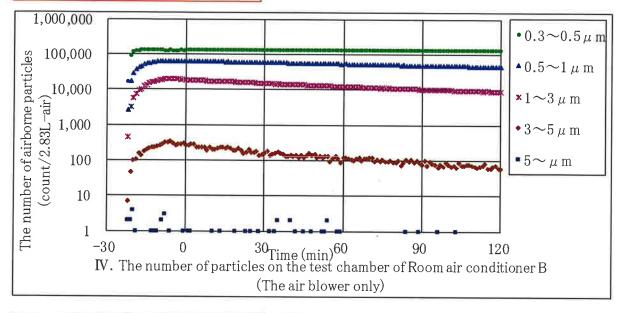
Fig 6. 25 m³ Test chamber (side view)

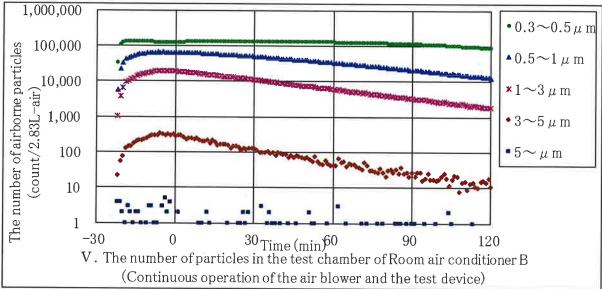


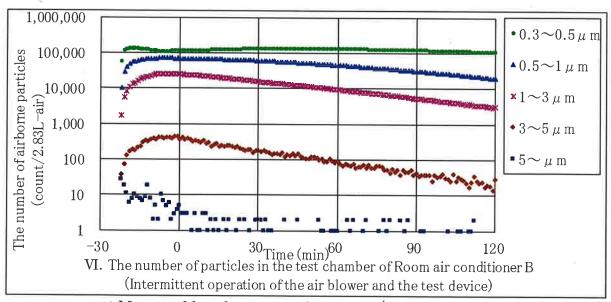




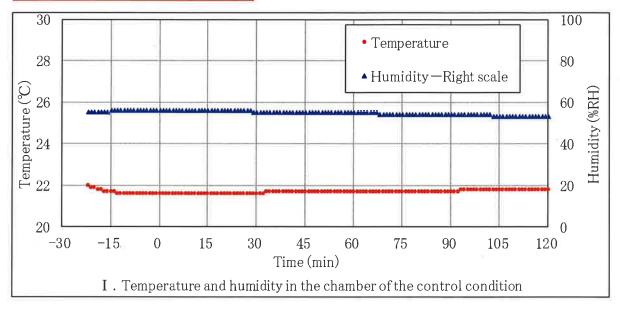
*Measured by a laser particle counter (MODEL 3886, Kanomax Japan)

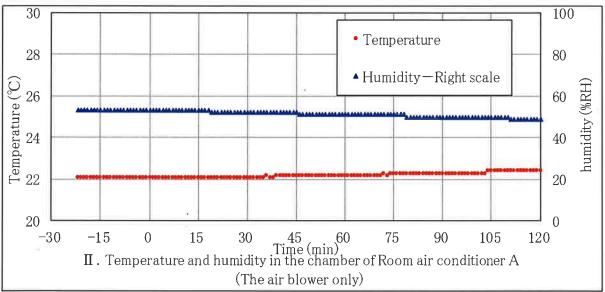


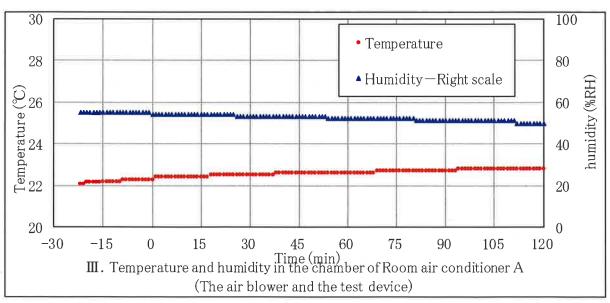




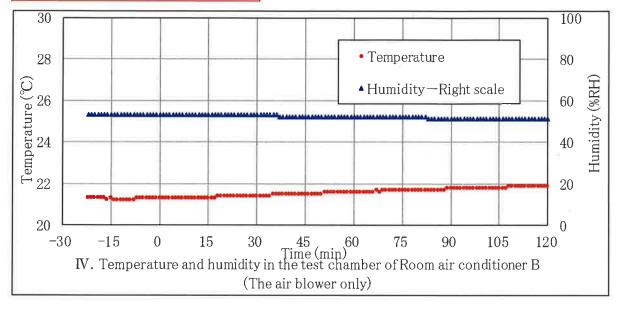
*Measured by a laser particle counter (MODEL 3886, Kanomax Japan)

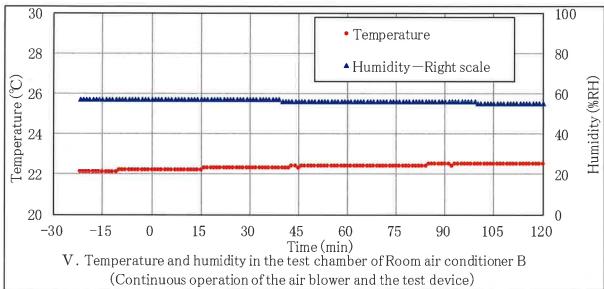


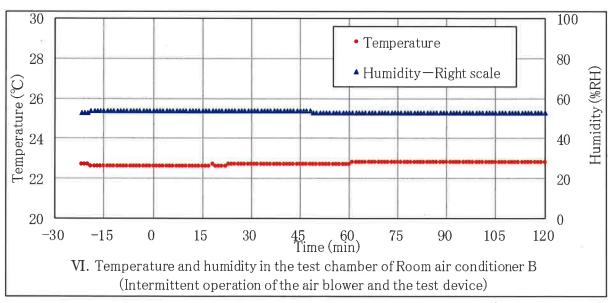




* Measured humidity in the chamber of the test device(TR-72Ui, T&D)







* Measured humidity in the chamber of the test device(TR-72Ui, T&D)

Attachment

Annex D: The evaluation test of removal efficiency for airborne virus.

Standard of household air cleaner, JEM 1467, The Japan Electrical Manufacturers' Association

Result

The changes of number of airborne phage or influenza virus are shown in Fig 1. The inclination of the approximate equation represents the changes of number of airborne phage or influenza virus per minute (log). The changes of the logarithmic values in this case mean the digit changes of the number of phage or influenza virus. Accordingly, the log reduction of "② test device running" at t min corrected with that of "① natural reduction" at t min is the log reduction of the number of phage or influenza virus with time.

The approximate equation is as follows;

Natural reduction :
$$y=-a_1x+b_1$$
 (D.1)

Test device running :
$$y=-a_2x+b_2$$
 (D.2)

y: Log (the number of the airborne phage or influenza virus (CFU / the values of the captured

air) x: The time of test device running (min)

The formulas Δy shows the logarithmic reduction value of airborne phage or influenza virus in the condition of natural reduction or running test device.

$$\Delta y = t(a_2 - a_1)$$
 (D.3)

One digit decrease meant 90% reduction, and 2 digit decrease meant 99% reduction.

$$\left(1 - \frac{1}{10^{\zeta}}\right) \times 100(\%) \tag{D.4}$$

 ζ : The decreasing number of digits

In calculating the logarithmic reduction value, the values from the extrapolation of the approximate equation must not be used and the actual measured values at each time must be used.

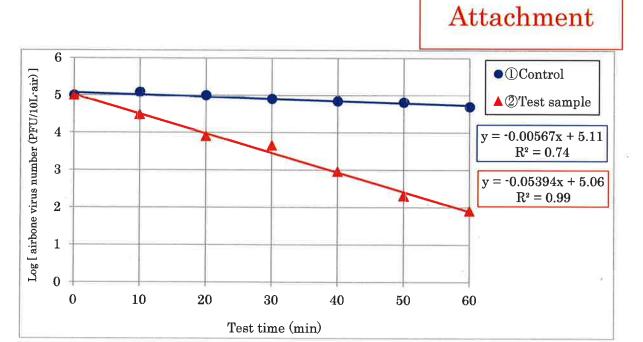


Fig D.1 Example of the results of removal efficiency evaluation test for airborne virus

Removal efficiency

When the logarithmic reduction value obtained from this examination is 2.0 or more, the air cleaner is considered as effective device for removing airborne virus.

To TOSHIBA CARRIER (THAILAND) CO., LTD.

Test Report

Evaluation test of the removal efficacy of the Room air conditioner on airborne fungal spores

(25 m³ space)

KRCES - Bio. Test Report No.2018_0356 February 15, 2019

Approved by: Toshihiro Al

Toshihiro Itoh, Chief Director

Kitasato Research Center for Environmental Science
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1. Test objective

Removal efficacy of the room air conditioners on airborne fungal spores in a 25 m³ test chamber was evaluated in this study. Evaluation method for removal efficacy was referring to Annex D, "The evaluation test for removal efficacy on airborne virus" in Japan Electrical Manufacturers' Association standard JEM 1467 "household air cleaner".

2. Client

Name: TOSHIBA CARRIER (THAILAND) CO., LTD.

Address: 144/9 Moo 5, Bangkadi, Industrial Park, Tivanon Road, Tambol Bangkadi, Amphur, Muang, Pathunthani 12000, Thailand

3. Test laboratory

Name: Kitasato Research Center for Environmental Science

Address: 1-15-1 Kitasato, Minami, Sagamihara, Kanagawa 252-0329, Japan

4. Test period

January 1, 2019 ~ January 23, 2019

5. Test sample

- 1) Room air conditioner A (Model No; RAS-35PKVPG-ND, Device; DSK9 with plasma) · · · Photo1
- 2) Room air conditioner B (Model No; RAS-13J2KCVRG-T, Device; BF1-13K with ionizer) • Photo2

6. Test condition

I. Natural reduction (control);

Time-dependent changes in fungal spores count were monitored after the test fungal spores suspension was splayed into the chamber where the test sample was turned off.

II. Room air conditioner A (The air blower only)

Time-dependent changes in fungal spores count were monitored after the test fungal spores suspension was sprayed into the chamber where only the air blower of the test sample was turned on. III. Room air conditioner A (The air blower and the test device)

Time-dependent changes in fungal spores count were monitored after the test fungal spores suspension was sprayed into the chamber where the sample (the air blower and the test device) was turned on.

IV. Room air conditioner B (The air blower only)

Time-dependent changes in fungal spores count were monitored after the test fungal spores suspension was sprayed into the chamber where only the air blower of the test sample was turned on.

V. Room air conditioner B (Continuous operation of the air blower and the test device)

Time-dependent changes in fungal spores count were monitored after the test fungal spores suspension was sprayed into the chamber where the sample (the air blower and the test device) was continuously operated for 30 seconds, and then turn off for 2 seconds.

VI. Room air conditioner B (Intermittent operation of the air blower and the test device)

Time-dependent changes in fungal spores count were monitored after the test fungal spores suspension was sprayed into the chamber where the sample (the air blower and the test device) was intermittently operated for 4 seconds, and then turn off for 6 seconds.

7. Test fungus

Penicillium citrinum NBRC 6352 (Spore), Size: approximately 3 µm

- 8. Reagents, apparatus
 - 1) Main reagents
 - · Potato Dextrose Agar (Nissui, hereinafter called PDA)
 - · Sodium chloride (Wako, special grade, for physiological saline)
 - · Sodium thiosulfate (Wako, 1st grade)
 - · Aerosol OT (Wako, for chemistry)
- 2) Devices and apparatus
 - Test chamber (25 m 3: 2.7×3.8×2.4 m, Amenity Technology)
 - · Circulation fan (BS-B-25, Yamazen)
 - · Laser particle counter (MODEL 3886, Kanomax Japan)
 - Thermo-hygrometer (TR-72Ui, T&D)

- · Nebulizer (Collison Nebulizer CN-31I、BGI)
- · Glass impinger (specially ordered, hereinafter called impinger)
- Membrane filter (ϕ 0.45 μ m, mixed cellulose ester, A045R047A, Advantec)
- · Incubator (MIR-153, MIR-553, Sanyo)

9. Method

1) Test procedure

The test system was shown in Photos 3, 4 and Figs 5, 6. The test sample, the circulation fan, the laser particle counter, and the thermo-hygrometer were set in the test chamber. Two holes were made on one of the side panels of the test chamber. The nebulizer for spraying fungal spores suspension was connected to one of the holes and the impinger for collecting airborne fungal spores was connected to the other hole.

According to the test procedure described in Table 6, the fungal spores suspension was sprayed with nebulizer for 15 minutes into the chamber while the circulation fan was operated. After 2 minute circulation of the air, the fungal spores aerosol was collected into the impinger (time 0). The aerosol was collected after 60 and 120 minutes after the test sample was turned on.

As a control, the same test was performed under the condition that the test sample was turned off according to the procedure described in Table 5.

2) Preparation of the test fungus

Cryopreserved test fungi were applied to PDA and incubated for 2 weeks at $27 \pm 2^{\circ}$ C. Spores were scraped off and suspended in sterile ion-exchanged water and filtrated through cotton wool. The fungal count was adjusted to 10^{8} CFU/mL for the test.

3) Spray of the fungal spores suspension

The test fungal spores suspension (10⁸ CFU/mL) was sprayed into the test chamber with the nebulizer for 15 minutes at the liquid flow rate of 0.2 mL/min. The air pressure from the compressor discharge was set at 1.5 kg/cm² and the air flow rate was set to 6.5 L/min.

4) Collection of airborne fungal spores

The air in the chamber was sampled at 10 L/min for 5 minutes (total 50 L) to the impinger containing 20 mL of sterilized saline with 0.015 % sodium thiosulfate to collect the airborne fungal spores (fungal spores aerosol).

5) Count of fungal spore numbers

Ten-fold dilutions of the each collected spore suspension were prepared with 0.005 % dioctyl sodium sulfosuccinate in sterilized saline. One mL of the each dilution or the original suspension was mixed with PDA medium to make an agar plate. Ten mL of the suspension of the collected fungal spore suspension in the impinger were filtered through the membrane filter. The resultant filter was transferred onto the surface of PDA medium. These medium were incubated at $27 \pm 2^{\circ}$ C for 1 week. After the incubation, colony numbers were counted and the numbers of fungal spores in 50 L of air were calculated.

6) Evaluation method for the removal efficacy

This test was carried out using Annex D of JEM 1467 as a reference. In JEM 1467, achieving 2.0-digit reduction in 90 minutes in the test space of $20\sim32$ m³ is required to conclude that the test sample is effective.

However, this test sample did not correspond to household air purifiers, and besides, the fungal spores were used instead of virus in this test. Therefore, the evaluation of the removal efficacy was performed using the method described below.

The approximate equation was calculated based on the time-dependent changes of airborne fungal spores (logarithmic representation) and the inclination of the approximate equation was obtained. This inclination represents an amount of change in the number of fungal spores per minute. Net inclination^{*1} was calculated by subtracting the inclination of control condition from that of test condition. Net LRV^{*2} and percent reduction^{*3} were calculated from the value of net inclination, and the removal efficacy of airborne fungal spores was determined based on net LRV.

The efficacy of the test sample was evaluated using the following formulae.

*1 Net inclination =

Inclination of test sample condition — Inclination of control condition $*_2$ Net LRV = -{ Net inclination \times Test time (min) }

**3 Percent reduction(%) =
$$\left(1 - \frac{1}{10^{\text{(Net LRV)}}}\right) \times 100 \text{ (%)}$$

10. Results

Fungal spore counts of the sprayed suspension was 6.8×10^8 CFU/mL. The numbers of airborne fungal spores at each measurement time were shown in Table 1 and Fig 1. Net LRV and percent of reduction were calculated from the number of airborne fungal spores at each measurement time, and were shown in Table 2 and Fig 2. Residual fungal spore ratios for the initial number at each time were shown in Table 3 and Fig 3. The residual fungal spore ratios for the control were shown in Table 4 and Fig 4.

In this test, net LRV (reduction rate) of the test samples for airborne fungal spores at 120 minutes were 1.30 (94%) for test condition II, 2.01 (99.0%) for the condition III, 1.20 (93%) for the condition IV, 2.30 (99.4%) for the condition V and 2.38 (99.5%) for the condition VI respectively.

11. Reference data

The number of airborne particles, temperature and humidity in the test chamber were shown as a reference data.

12. Comment

This test was referring to the method of JEM 1467. In attached document Annex D, "The evaluation test for removal efficacy on airborne virus" in Japan Electrical Manufacturers' Association standard JEM 1467, achieving 2.0-digit reduction in 90 minutes in the test space of 20~32 m³ is required to conclude that the test sample is effective.

In the test conditions I and IV, the test sample showed certain degree of reduction effect but could not achieved 2.0 log reduction value even after being operated for 120min. On the other hand, in the test conditions III, IV and V, the log reduction value was more than 2-dight, and removed efficacy of the test sample against airborne fungi was recognized.

The two major differences between this standard and this test must be considered, as follows.

- 1) The test samples in this test were not designed as air purifiers but air conditioners.
- 2) The target microorganisms used in this test were bacteria instead of viruses.

Table 1. The number of airborne fungal spores at each measurement time (Unit: CFU/100 L-air)

Test condition		Time (min)			
rest condition	0	60	120		
I. Control condition	330,000	190,000	120,000		
II . Room air conditioner A	380,000	37,000	6,800		
(The air blower only)	380,000	37,000	0,000		
III. Room air conditioner A	400,000	12,000	1 400		
(The air blower and the test device)	400,000	12,000	1,400		
IV. Room air conditioner B	410,000	42,000	0.200		
(The air blower only)	410,000	43,000	9,300		
V. Room air conditioner B	940,000	10.000	1 500		
(Continuous operation of the air blower and the test device)	840,000	19,000	1,500		
VI. Room air conditioner B	1 000 000	21.000	1.500		
(Intermittent operation of the air blower and the test device)	1,000,000	31,000	1,500		

Test sample:

- 1) Room air conditioner A (Model No; RAS-35PKVPG-ND, Device; DSK9 with plasma)
- 2) Room air conditioner B (Model No; RAS-13J2KCVRG-T, Device; BF1-13K with ionizer)

Test fungus: Penicillium citrinum NBRC 6352 (Spore)

Test space: 25 m³

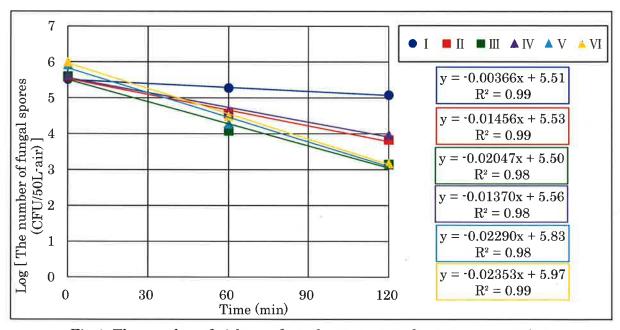


Fig 1. The number of airborne fungal spores at each measurement time

Table 2. Net LRV and percent reduction at each measurement tim	Table 2.	Net LRV an	percent reduction	at each measurement time
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Test condition	Inclination	Net	7	Time (min)
Test condition	Inclination	inclination	0	60	120
I. Control condition	-0.00366				
II. Room air conditioner A	-0.01456	-0.01090	0.00	0.65	1.30
(The air blower only)			(0%)	(77%)	(94%)
III. Room air conditioner A	-0.02047	-0.01681	0.00	1.00	2.01
(The air blower and the test device)			(0%)	(90%)	(99.0%)
IV. Room air conditioner B	-0.01370	-0.01004	0.00	0.60	1.20
(The air blower only)			(0%)	(74%)	(93%)
V. Room air conditioner B			0.00	1.15	2.30
(Continuous operation of the air blower and the test device)	-0.02290	-0.01924	(0%)	(92%)	(99.4%)
VI. Room air conditioner B		70	0.00	1.19	2.38
(Intermittent operation of the air blower and the test device)	-0.02353	-0.01987	(0%)	(93%)	(99.5%)

 $Net\ inclination\ =\ Inclination\ of\ test\ condition\ -\ Inclination\ of\ control\ condition$

Net LRV = -{ Net inclination \times Test time (min) }

Percent reduction (%) =
$$\left(1 - \frac{1}{10^{\text{(Net LRV)}}}\right) \times 100 \text{ (%)}$$

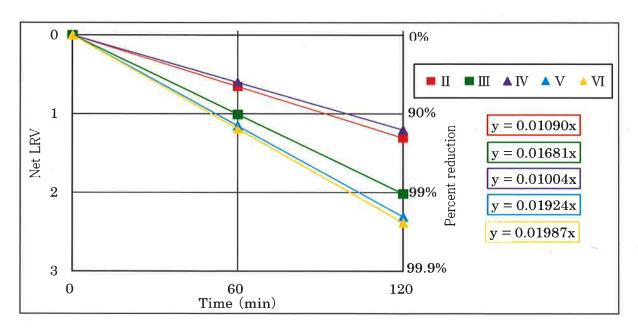


Fig 2. Net LRV and percent reduction at each measurement time

Table 3.	Residual fungal	spore ratio for the	he initial n	umber (%)	at each time

Test condition		Time (min)						
		20	40	60	80	100	120	
I. Control condition	100	84	71	60	50	43	36	
II. Room air conditioner A (The air blower only)	100	51	26	13	6.8	3.4	1.7	
III. Room air conditioner A (The air blower and the test device)		38	15	5.9	2.3	0.8	0.3	
IV. Room air conditioner B (The air blower only)		53	28	15	8.0	4.2	2.2	
V. Room air conditioner B (Continuous operation of the air blower and the test device)		34	12	4.2	1.4	0.5	0.1	
VI. Room air conditioner B (Intermittent operation of the air blower and the test device)	100	33	11	3.8	1.3	0.4	0.1	

*Using slope of approximate expression as shown in Fig1 (number of airborne fungal spores which change per minute, log value) the residual ratio was calculated according to equation ① and ②.

① Log reduction value = | Inclination | ×Time (min)

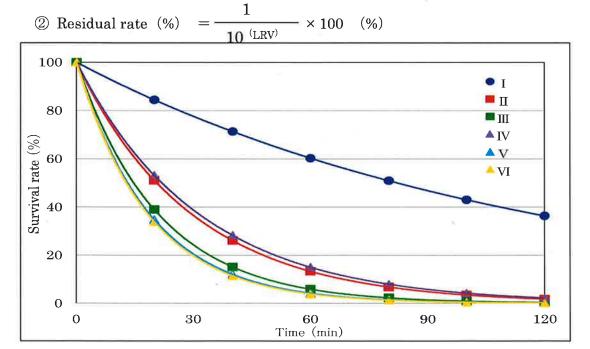


Fig 3. Residual fungal spore ratio for the initial number (%) at each time

Table 4.	The residual	fungal spore	ratios with	the sample	for the contro	l at each time
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Test condition		Time (min)						
		20	40	60	80	100	120	
I. Control condition		100	100	100	100	100	100	
II . Room air conditioner A (The air blower only)		60	36	22	13	8.1	4.9	
III. Room air conditioner A (The air blower and the test device)		46	21	9.8	4.5	2.0	0.9	
IV. Room air conditioner B (The air blower only)		62	39	24	15	9.9	6.2	
V. Room air conditioner B (Continuous operation of the air blower and the test device)		41	16	7.0	2.8	1.1	0.4	
VI. Room air conditioner B (Intermittent operation of the air blower and the test device)		40	16	6.4	2.5	1.0	0.4	

*Using slope of approximate expression as shown in Fig1 (number of airborne fungal spores which change per minute, log value) the residual ratio was calculated according to equation ①,② and ③.

- ① Net inclination = Inclination of test condition Inclination of control condition
- ② LRV= | Net Inclination | ×Time (min)

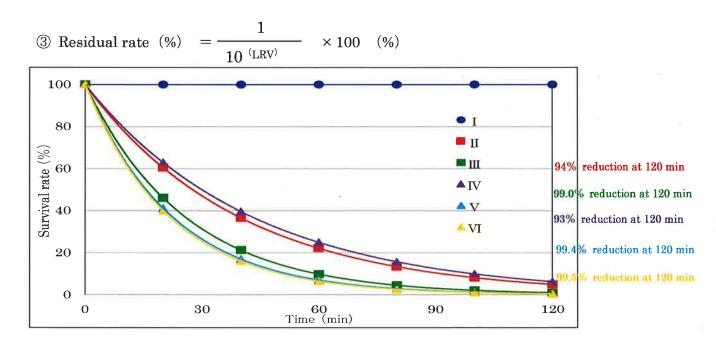


Fig 4. The residual fungal spore ratios with the sample for the control at each time

Table 5. Test procedure (Control condition I)

Test condition	Equipment	Time (min)					
Test condition	Equipment	() 6	80	120		
To make homogeneous air in the chamber	Circulation fan				-		
Spray of fungal spores	Nebulizer	15min Disper	sing bacteria for 2	min			
Collect airborne fungal spores	Impinger	5min		5min 10L/min	5min 10L/min		

Table 6. Test procedure (Test condition $II \sim VI$)

Test condition	Equipment	Time (min)				
Test condition	Equipment	0	60	120		
To make homogeneous air in the chamber	Circulation fan			-		
Spray of fungal spores	Nebulizer	15min Dispersing b	acteria for 2 min			
Test sample	Air blower			-		
rest sample	Test device					
Collect airborne fungal spores	Impinger	5min 10L/min	5min 10L/min	5min 10L/min		



Photo 1. Room air conditioner A (Model No ; RAS-35PKVPG-ND)



Photo 2. Room air conditioner B (Model No; RAS-13J2KCVRG-T)



Photo 3. 25 m³ Test chamber (Room air conditioner A)



Photo 4. 25 m³ Test chamber (Room air conditioner B)

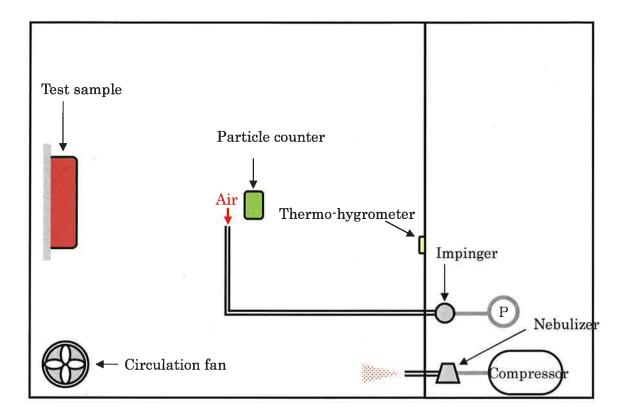


Fig 5. 25 m³ Test chamber (top view)

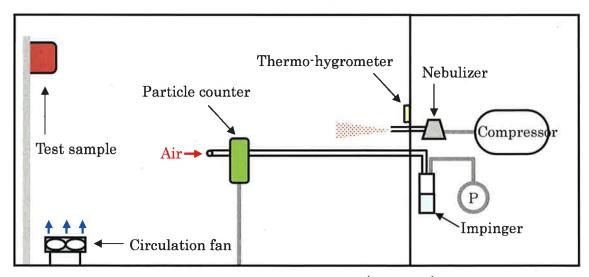
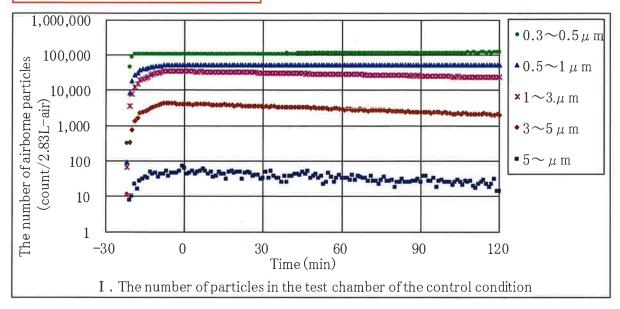
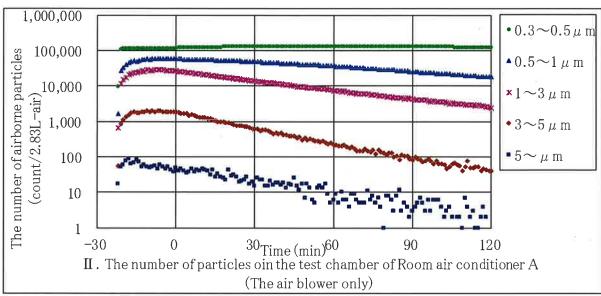
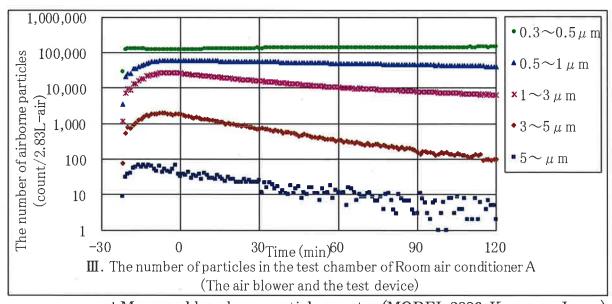


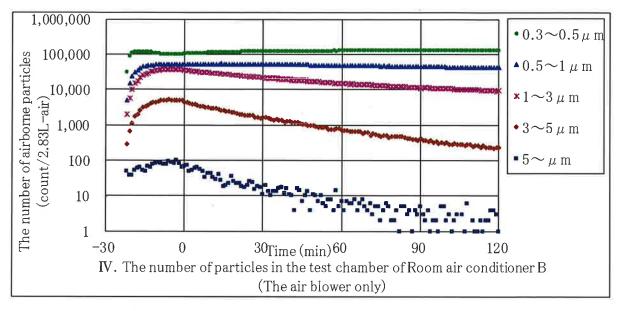
Fig 6. 25 m³ Test chamber (side view)

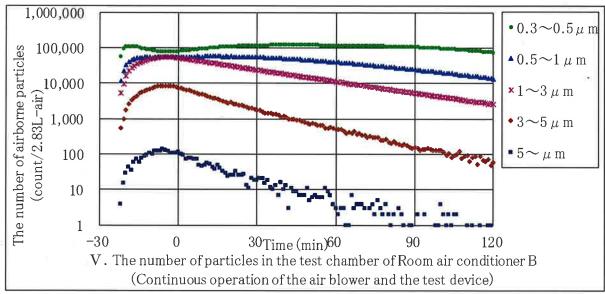


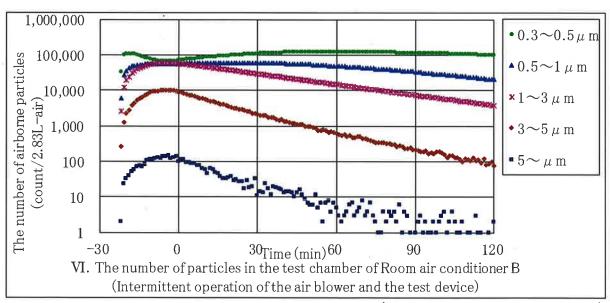




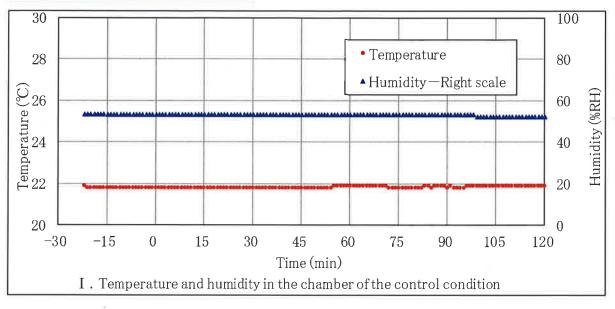
*Measured by a laser particle counter (MODEL 3886, Kanomax Japan)

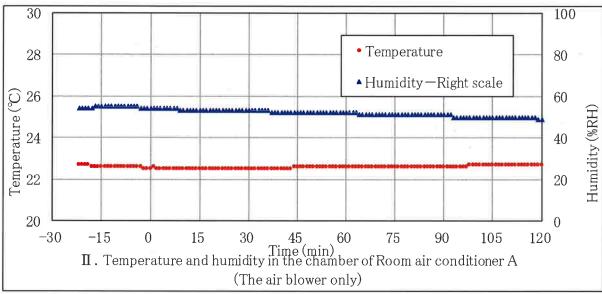


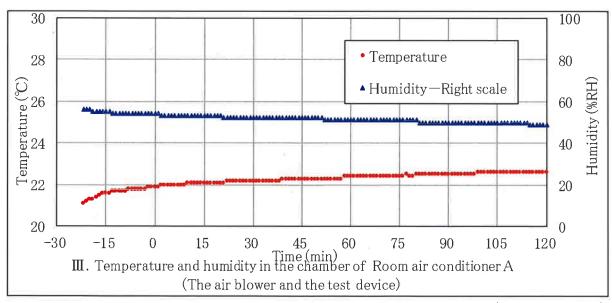




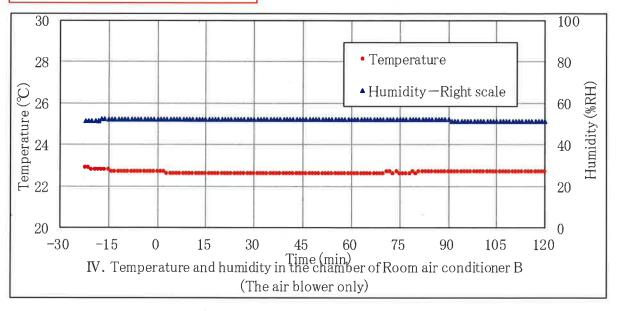
*Measured by a laser particle counter (MODEL 3886, Kanomax Japan)

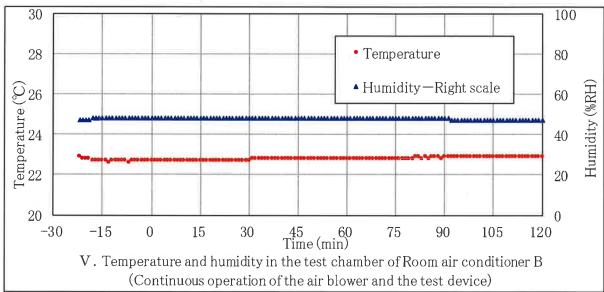


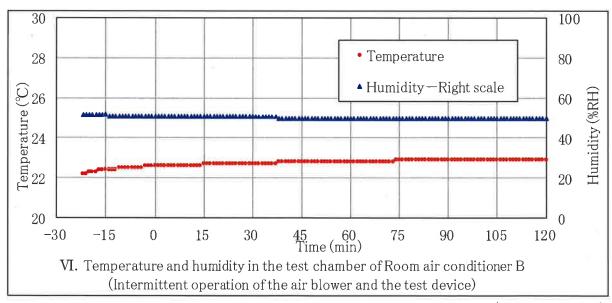




* Measured humidity in the chamber of the test device(TR-72Ui, T&D)







* Measured humidity in the chamber of the test device(TR-72Ui, T&D)

Annex D: The evaluation test of removal efficiency for airborne virus.

Standard of household air cleaner, JEM 1467, The Japan Electrical Manufacturers' Association

Result

The changes of number of airborne phage or influenza virus are shown in Fig 1. The inclination of the approximate equation represents the changes of number of airborne phage or influenza virus per minute (log). The changes of the logarithmic values in this case mean the digit changes of the number of phage or influenza virus. Accordingly, the log reduction of "② test device running" at t min corrected with that of "① natural reduction" at t min is the log reduction of the number of phage or influenza virus with time.

The approximate equation is as follows;

Natural reduction :
$$y=-a_1x+b_1$$
 (D.1)

Test device running:
$$y=-a_2x+b_2$$
(D.2)

y: Log (the number of the airborne phage or influenza virus (CFU / the values of the captured

air)) x: The time of test device running (min))

The formulas Δy shows the logarithmic reduction value of airborne phage or influenza virus in the condition of natural reduction or running test device.

$$\Delta y = t(a_2 - a_1)$$
 (D.3)

One digit decrease meant 90% reduction, and 2 digit decrease meant 99% reduction.

$$\left(1 - \frac{1}{10^{\zeta}}\right) \times 100(\%)$$
 (D.4)

 ζ : The decreasing number of digits

In calculating the logarithmic reduction value, the values from the extrapolation of the approximate equation must not be used and the actual measured values at each time must be used.

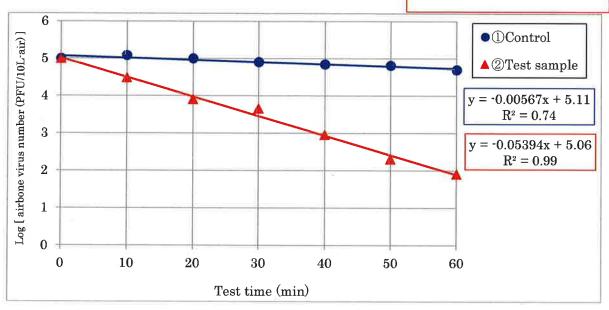


Fig D.1 Example of the results of removal efficiency evaluation test for airborne virus

Removal efficiency

When the logarithmic reduction value obtained from this examination is 2.0 or more, the air cleaner is considered as effective device for removing airborne virus.

1. Test objective

Removal efficacy of the room air conditioners on airborne bacteria in a 25 m³ test chamber was evaluated in this study. Evaluation method for removal efficacy was referring to Annex D, "The evaluation test for removal efficacy on airborne virus" in Japan Electrical Manufacturers' Association standard JEM 1467 "household air cleaner".

2. Client

Name: TOSHIBA CARRIER (THAILAND) CO., LTD.

Address: 144/9 Moo 5, Bangkadi, Industrial Park, Tivanon Road, Tambol Bangkadi, Amphur, Muang, Pathunthani 12000, Thailand

3. Test laboratory

Name: Kitasato Research Center for Environmental Science

Address: 1-15-1 Kitasato, Minami, Sagamihara, Kanagawa 252-0329, Japan

4. Test period

January 15, 2019 ~ January 18, 2019

5. Test sample

- 1) Room air conditioner A (Model No; RAS-35PKVPG-ND, Device; DSK9 with plasma) · · · Photo1
- 2) Room air conditioner B (Model No; RAS-13J2KCVRG-T, Device; BF1-13K with ionizer) · · · Photo2

6. Test condition

I. Natural reduction (control);

Time-dependent changes in bacteria count were monitored after the test bacterial suspension was splayed into the chamber where the test sample was turned off.

II. Room air conditioner A (The air blower only)

Time-dependent changes in bacteria count were monitored after the test bacterial suspension was sprayed into the chamber where only the air blower of the test sample was turned on. III. Room air conditioner A (The air blower and the test device)

Time-dependent changes in bacteria count were monitored after the test bacterial suspension was sprayed into the chamber where the sample (the air blower and the test device) was turned on.

IV. Room air conditioner B (The air blower only)

Time-dependent changes in bacteria count were monitored after the test bacterial suspension was sprayed into the chamber where only the air blower of the test sample was turned on.

V. Room air conditioner B (Continuous operation of the air blower and the test device)

Time-dependent changes in bacteria count were monitored after the test bacterial suspension was sprayed into the chamber where the sample (the air blower and the test device) was continuously operated for 30 seconds, and then turn off for 2 seconds.

VI. Room air conditioner B (Intermittent operation of the air blower and the test device)

Time-dependent changes in bacteria count were monitored after the test bacterial suspension was sprayed into the chamber where the sample (the air blower and the test device) was intermittently operated for 4 seconds, and then turn off for 6 seconds.

7. Test bacteria

Staphylococcus aureus NBRC 12732, Size: approximately 0.5 µm

- 8. Reagents, apparatus
 - 1) Main reagents
 - Tryptic Soy Agar (Difco, hereinafter called TSA)
 - · Sodium chloride (Wako, special grade, for physiological saline)
 - · Sodium thiosulfate (Wako, 1st grade)
- 2) Devices and apparatus
 - Test chamber (25 m ³: 2.7×3.8×2.4 m, Amenity Technology)
 - · Circulation fan (BS-B-25, Yamazen)
 - · Laser particle counter (MODEL 3886, Kanomax Japan)
 - Thermo-hygrometer (TR-72Ui, T&D)
 - Nebulizer (Collison Nebulizer CN-31I、BGI)

- · Glass impinger (specially ordered, hereinafter called impinger)
- Membrane filter (ϕ 0.45 μ m, mixed cellulose ester, A045R047A, Advantec)
- · Incubator (MIR-153, MIR-553, Sanyo)

9. Method

1) Test procedure

The test system was shown in Photos 3, 4 and Figs 5, 6. The test sample, the circulation fan, the laser particle counter, and the thermo-hygrometer were set in the test chamber. Two holes were made on one of the side panels of the test chamber. The nebulizer for spraying bacterial suspension was connected to one of the holes and the impinger for collecting airborne bacteria was connected to the other hole.

According to the test procedure described in Table 6, the bacterial suspension was sprayed with nebulizer for 15 minutes into the chamber while the circulation fan was operated. After 2 minute circulation of the air, the bacterial aerosol was collected into the impinger (time 0). The aerosol was collected after 60 and 120 minutes after the test sample was turned on.

As a control, the same test was performed under the condition that the test sample was turned off according to the procedure described in Table 5.

2) Preparation of test bacteria

Cryopreserved test bacteria were pre-cultured and then sub-cultured for 21 hours at 36 ± 2 °C on TSA. Colonies formed on TSA were scraped off and suspended in sterilized ion-exchange water. Bacterial count of the suspension was adjusted to about 10^9 CFU/mL for the test.

3) Spray of the bacterial suspension

The test bacterial suspension (10° CFU/mL) was sprayed into the test chamber with the nebulizer for 15 minutes at the liquid rate of 0.2 mL/min. The air pressure from the compressor discharge was set at 1.5 kg/cm² and the air flow rate was set to 7.0 L/min.

4) Collection of airborne bacteria

The air in the chamber was sampled at 10 L/min for 5 minutes (total 50 L) to the impinger containing 20 mL of sterilized saline with 0.015 % sodium thiosulfate to collect the airborne bacteria (bacterial aerosol).

5) Count of bacterial numbers

Ten-fold dilutions of the each collected bacterial suspension were prepared with saline. One mL of the each dilution or the original suspension was mixed with TSA medium to make an agar plate. Ten mL of the suspension of the collected bacterial suspension in the impinger were filtered through the membrane filter. The resultant filter was transferred onto the surface of TSA medium. These media were incubated at 36 ± 2 °C for 43 hours. After the incubation, colony numbers were counted and the numbers of bacteria in 50 L of air were calculated.

6) Evaluation method for the removal efficacy

This test was carried out using Annex D of JEM 1467 as a reference. In JEM 1467, achieving 2.0-digit reduction in 90 minutes in the test space of $20\sim32$ m³ is required to conclude that the test sample is effective.

However, this test sample did not correspond to household air purifiers, and besides, the bacteria were used instead of virus in this test. Therefore, the evaluation of the removal efficacy was performed using the method described below.

The approximate equation was calculated based on the time-dependent changes of airborne bacteria (logarithmic representation) and the inclination of the approximate equation was obtained. This inclination represents an amount of change in the number of bacteria per minute. Net inclination*1 was calculated by subtracting the inclination of control condition from that of test condition. Net LRV* 2 and percent reduction*3 were calculated from the value of net inclination, and the removal efficacy of airborne bacteria was determined based on net LRV.

The efficacy of the test sample was evaluated using the following formulae.

*1 Net inclination =

Inclination of test sample condition — Inclination of control condition $*_2$ Net LRV = $-\{$ Net inclination \times Test time (min) $\}$

**3 Percent reduction(%) =
$$\left[1 - \frac{1}{10^{\text{(Net LRV)}}}\right] \times 100 \text{ (%)}$$

10. Results

Bacterial count of the sprayed suspension was 2.5×10^9 CFU/mL. The numbers of airborne bacteria at each measurement time were shown in Table 1 and Fig 1. Net LRV and percent of reduction were calculated from the number of airborne bacteria at each measurement time, and were shown in Table 2 and Fig 2. Residual bacteria ratios for the initial number at each time were shown in Table 3 and Fig 3. The residual bacteria ratios for the control were shown in Table 4 and Fig 4.

In this test, net LRV (reduction rate) of the test samples for airborne bacteria at 120 minutes were 0.17 (32%) for test condition II, 0.73 (81%) for the condition III, 0.12 (24%) for the condition IV, 1.35 (95%) for the condition V and 0.90 (87%) for the condition VI respectively.

11. Reference data

The number of airborne particles, temperature and humidity in the test chamber were shown as a reference data.

12. Comment

This test was referring to the method of JEM 1467. In attached document Annex D, "The evaluation test for removal efficacy on airborne virus" in Japan Electrical Manufacturers' Association standard JEM 1467, achieving 2.0-digit reduction in 90 minutes in the test space of 20~32 m³ is required to conclude that the test sample is effective.

In this study, the test sample showed certain degree of reduction effect but could not achieved 2.0 log reduction value even after being operated for 120min.

The two major differences between this standard and this test must be considered, as follows.

- 1) The test samples in this test were not designed as air purifiers but air conditioners.
- 2) The target microorganisms used in this test were bacteria instead of viruses.

Table 1. The number of airborne bacteria at each measurement time (Unit : CFU/50 L-air)

Test condition	Time (min)				
Test condition	0	60	120		
I. Control condition	1,900,000	770,000	520,000		
II. Room air conditioner A	2 500 000	000 000	460,000		
(The air blower only)	2,500,000	990,000	460,000		
III. Room air conditioner A	2 900 000	460,000	100.000		
(The air blower and the test device)	3,800,000	460,000	190,000		
IV. Room air conditioner B	1,500,000	540,000	310,000		
(The air blower only)	1,500,000	340,000	310,000		
V. Room air conditioner B	2 700 000	270,000	22.000		
(Continuous operation of the air blower and the test device)	2,700,000	270,000	33,000		
VI. Room air conditioner B	1,600,000	250,000	55,000		
(Intermittent operation of the air blower and the test device)	1,000,000	250,000	55,000		

Test sample:

- 1) Room air conditioner A (Model No; RAS-35PKVPG-ND, Device; DSK9 with plasma)
- 2) Room air conditioner B (Model No; RAS-13J2KCVRG-T, Device; BF1-13K with ionizer)

Test bacteria: Staphylococcus aureus NBRC 12732

Test space: 25 m³

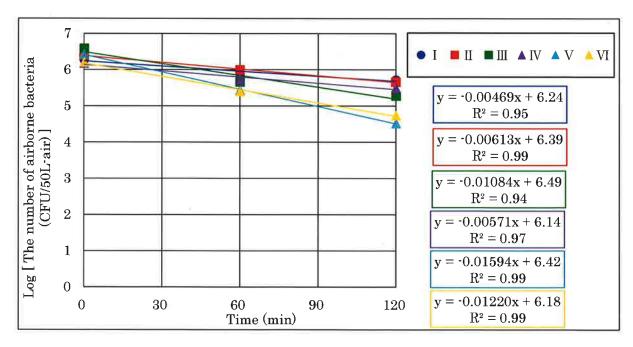


Fig 1. The number of airborne bacteria at each measurement time

Table 2. Net Lity and bereent reduction at each measurement to	Table 2.	e 2. Net LRV and	percent reduction at each	measurement time
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That and liting	In alimetica	Net	Time (min)			
Test condition	Inclination	inclination	0	60	120	
I. Control condition	-0.00469					
II. Room air conditioner A	-0.00613	-0.00144	0.00	0.08	0.17	
(The air blower only)			(0%)	(16%)	(32%)	
III. Room air conditioner A	-0.01084 -0.00615		0.00	0.36	0.73	
(The air blower and the test device			(0%)	(56%)	(81%)	
IV. Room air conditioner B	-0.00571	-0.00102	0.00	0.06	0.12	
(The air blower only)			(0%)	(12%)	(24%)	
V. Room air conditioner B	0.01504	0.01105	0.00	0.67	1.35	
(Continuous operation of the air blower and the test device)	-0.01594	-0.01125	(0%)	(78%)	(95%)	
VI. Room air conditioner B			0.00	0.45	0.90	
(Intermittent operation of the air blower and the test device)	-0.01220	-0.00751	(0%)	(64%)	(87%)	

Net inclination = Inclination of test condition - Inclination of control condition

Net LRV = $-\{$ Net inclination \times Test time (min) $\}$

Percent reduction (%) =
$$\left(1 - \frac{1}{10^{\text{(Net LRV)}}}\right) \times 100 \text{ (%)}$$

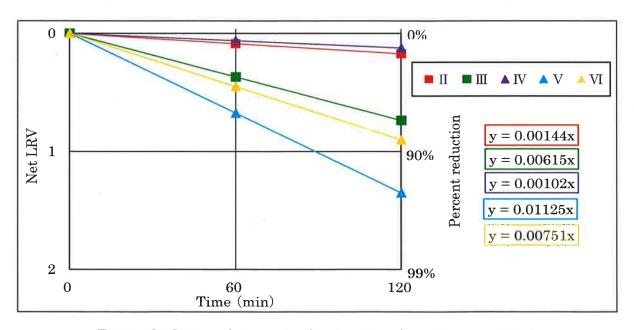


Fig 2. Net LRV and percent reduction at each measurement time

Table 3.	Residual	bacterial	ratio f	for the	initial	number	(%)	at each time

Test condition		Time (min)						
		20	40	60	80	100	120	
I. Control condition	100	80	64	52	42	33	27	
II. Room air conditioner A (The air blower only)	100	75	56	42	32	24	18	
III. Room air conditioner A (The air blower and the test device)	100	60	36	22	13	8.2	5.0	
IV. Room air conditioner B (The air blower only)	100	76	59	45	34	26	20	
V. Room air conditioner B (Continuous operation of the air blower and the test device)	100	47	23	11	5.3	2.5	1.2	
VI. Room air conditioner B (Intermittent operation of the air blower and the test device)	100	57	32	18	10	6.0	3.4	

**Using slope of approximate expression as shown in Fig1 (number of airborne bacteria which change per minute, log value) the residual ratio was calculated according to equation ① and ②.

① Log reduction value = | Inclination | ×Time (min)

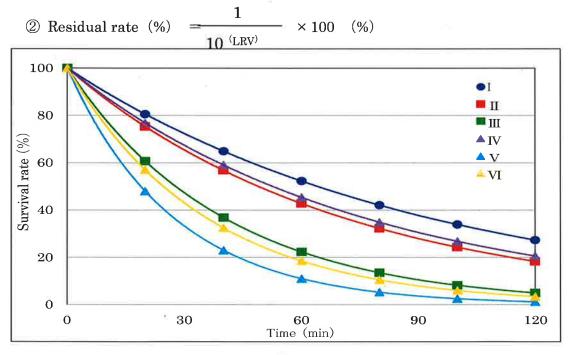


Fig 3. Residual bacterial ratio for the initial number (%) at each time

Table 4. The residual bacteria ratios with the sample for the control at each time

Test condition		Time (min)							
1 est condition	0	20	40	60	80	100	120		
I. Control condition	100	100	100	100	100	100	100		
II . Room air conditioner A (The air blower only)	100	93	87	81	76	71	67		
III. Room air conditioner A (The air blower and the test device)	100	75	56	42	32	24	18		
IV. Room air conditioner B (The air blower only)	100	95	91	86	82	79	75		
V. Room air conditioner B (Continuous operation of the air blower and the test device)	100	59	35	21	12	7.4	4.4		
VI. Room air conditioner B (Intermittent operation of the air blower and the test device)	100	70	50	35	25	17	12		

*Using slope of approximate expression as shown in Fig1 (number of airborne bacteria which change per minute, log value) the residual ratio was calculated according to equation ①,② and ③.

- ① Net inclination = Inclination of test condition Inclination of control condition
- ② LRV= | Net Inclination | ×Time (min)

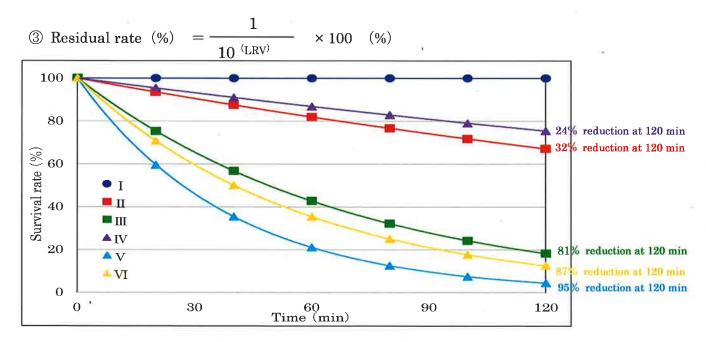


Fig 4. The residual bacteria ratios with the sample for the control at each time

Table 5. Test procedure (Control condition I)

Test condition	Equipment	Time (min)					
Test condition	Equipment	(0 60		.20		
To make homogeneous air in the chamber	Circulation fan				-		
Spray of bacteria	Nebulizer	15min Disper	sing bacteria for 2	min			
Collect airborne bacteria	Impinger	5min		5min 10L/min	5min 10L/min		

Table 6. Test procedure (Test condition $II \sim VI$)

Test condition	Equipment		Time (min)	
rest condition	Equipment	0	60	120
To make homogeneous air in the chamber	Circulation fan			
Spray of bacteria	Nebulizer	15min Dispersing b	acteria for 2 min	
Test semale	Air blower			-
Test sample	Test device			
Collect airborne bacteria	Impinger	5min 10L/min	5min 10L/min	5min



Photo 1. Room air conditioner A (Model No ; RAS-35PKVPG-ND)



Photo 2. Room air conditioner B (Model No ; RAS-13J2KCVRG-T)



Photo 3. 25 m³ Test chamber (Room air conditioner A)



Photo 4. 25 m³ Test chamber (Room air conditioner B)

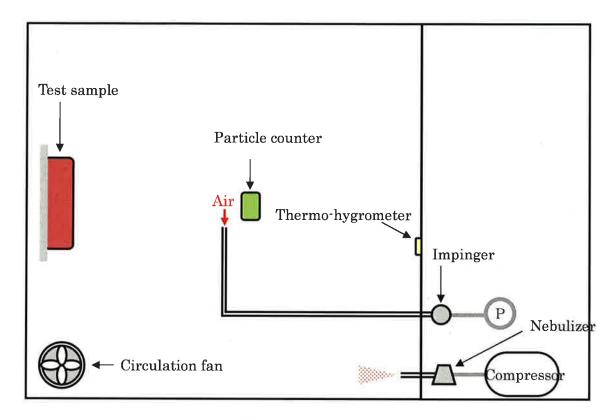


Fig 5. 25 m³ Test chamber (top view)

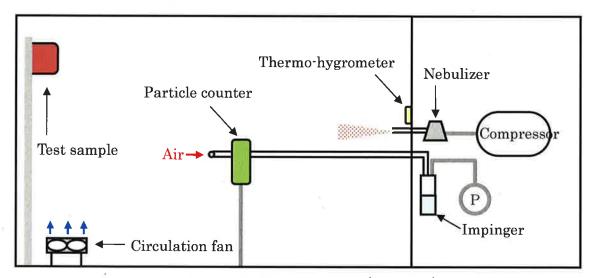
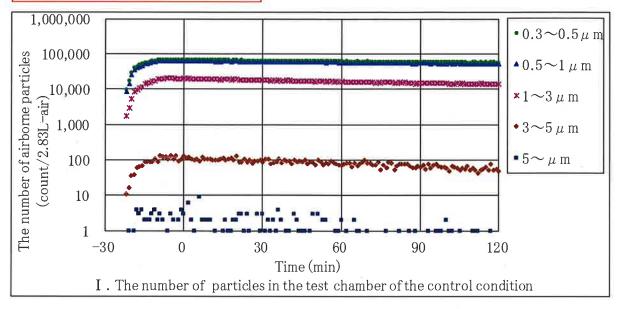
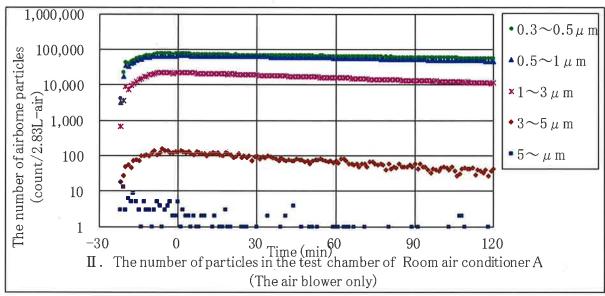
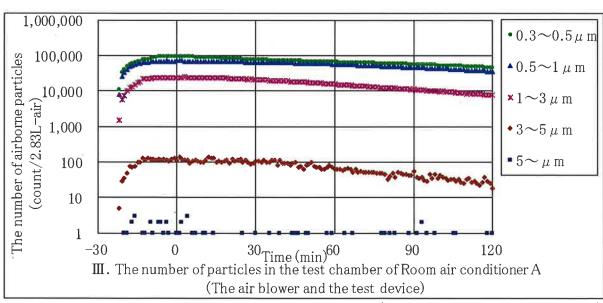


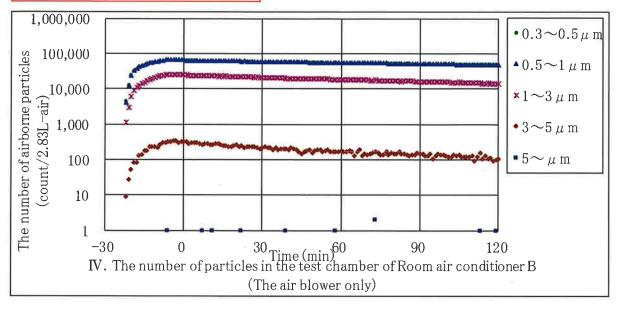
Fig 6. 25 m³ Test chamber (side view)

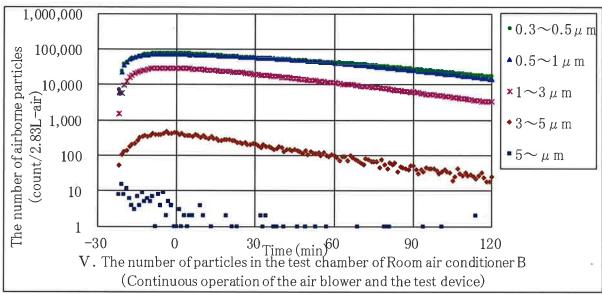


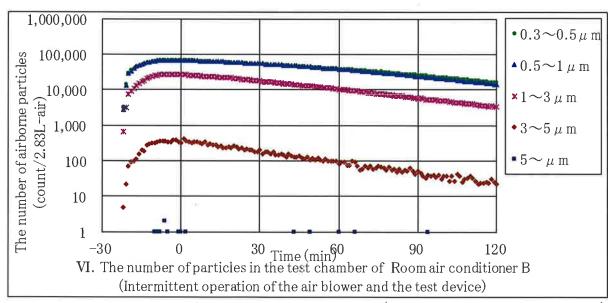




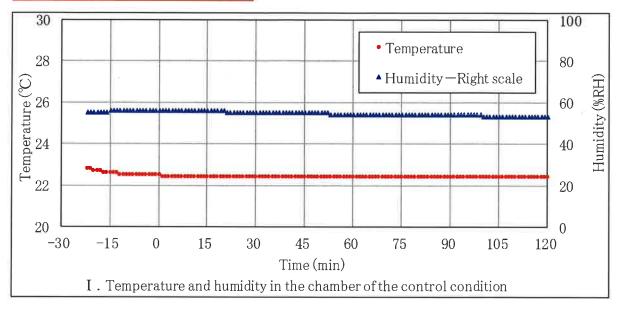
*Measured by a laser particle counter (MODEL 3886, Kanomax Japan)

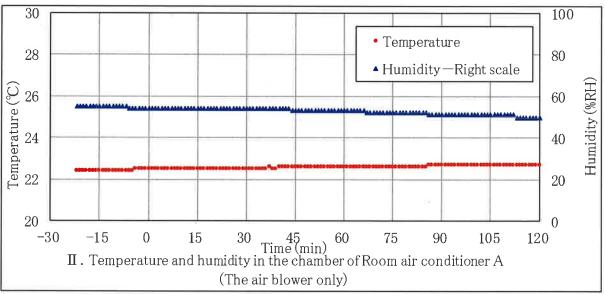


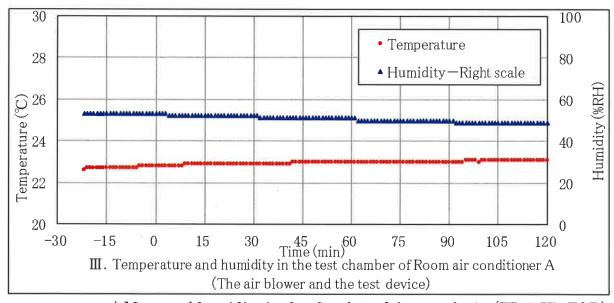




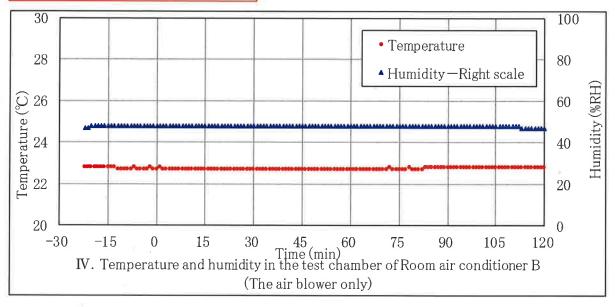
*Measured by a laser particle counter (MODEL 3886, Kanomax Japan)

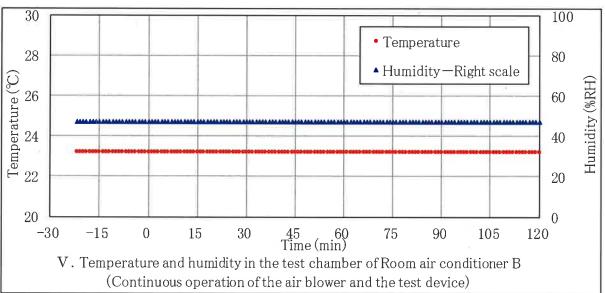


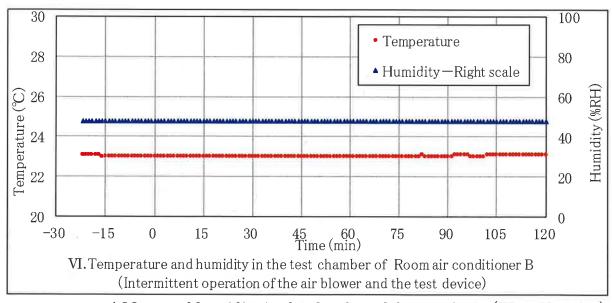




* Measured humidity in the chamber of the test device(TR-72Ui, T&D)







* Measured humidity in the chamber of the test device(TR-72Ui, T&D)

Annex D: The evaluation test of removal efficiency for airborne virus.

Standard of household air cleaner, JEM 1467, The Japan Electrical Manufacturers' Association

Result

The changes of number of airborne phage or influenza virus are shown in Fig 1. The inclination of the approximate equation represents the changes of number of airborne phage or influenza virus per minute (log). The changes of the logarithmic values in this case mean the digit changes of the number of phage or influenza virus. Accordingly, the log reduction of "② test device running" at t min corrected with that of "① natural reduction" at t min is the log reduction of the number of phage or influenza virus with time.

The approximate equation is as follows;

Natural reduction :
$$y=-a_1x+b_1$$
 (D.1)

Test device running:
$$y=-a_2x+b_2$$
 (D.2)

y: Log (the number of the airborne phage or influenza virus (CFU / the values of the captured air)) x: The time of test device running (min))

The formulas Δy shows the logarithmic reduction value of airborne phage or influenza virus in the condition of natural reduction or running test device.

$$\Delta y = t(a_2 - a_1)$$
 (D.3)

One digit decrease meant 90% reduction, and 2 digit decrease meant 99% reduction.

$$\left(1 - \frac{1}{10^{\zeta}}\right) \times 100(\%)$$
(D.4)

 ζ : The decreasing number of digits

In calculating the logarithmic reduction value, the values from the extrapolation of the approximate equation must not be used and the actual measured values at each time must be used.

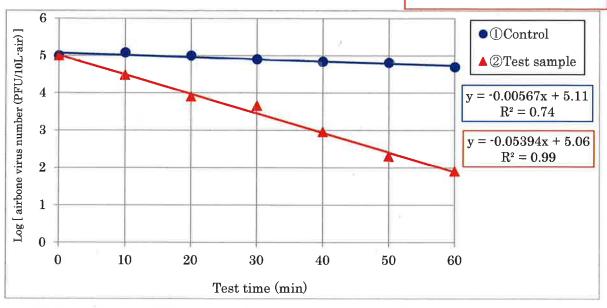


Fig D.1 Example of the results of removal efficiency evaluation test for airborne virus

Removal efficiency

When the logarithmic reduction value obtained from this examination is 2.0 or more, the air cleaner is considered as effective device for removing airborne virus.